

Viral Transport Media Collection Guidelines

General Collection Information

1. Label and Transport Instructions

- **Assure proper labeling of specimen. Standard labeling requirements include:**
 1. Patient's full legal name – complete first and last names correctly spelled
 2. Test Requisition Barcode, Medical Record Number or Date of Birth
 3. Date & time of collections (documented on sample or requisition)
 4. Name of collector (documented on sample or requisition)
 5. Source of specimen
- These collection guidelines serve only as general instructions. Only trained personnel should perform these procedures.
- For available testing, and further information, please refer to **Nationwide Children's Laboratory Guide to Services**
- Viral collection transport media is used to transport all types of swab specimens for viral, Chlamydial and Mycoplasma, PCR testing, and Respiratory Infection Array (FARVPP).
- Prior to sample collection, check that the viral transport medium: (1) has not passed the expiration date, (2) shows no signs of leakage, (3) has not changed color, or (4) shows no other signs of deterioration.
- Store viral transport media refrigerated or room temperature (2° C – 30° C) according to the temperature printed on vial label AND until printed expiration date. If the vial label does not have the temperature printed on the label, please call Client Services at (614) 722-5477.
- Once the sample has been collected in the media, the sample **MUST** be refrigerated and transported on wet ice or cold-pack.

2. Collection Kit Contents

- Sterile polyester- tipped swabs (enclosed in paper envelope) or sterile flocked swabs (enclosed in paper envelope)
- Viral transport media

3. General Swab Collection Instructions

- Apply appropriate but significant pressure to the body site to be tested while rotating the swab.
- This ensures that adequate cells (viruses and chlamydial species are intracellular) are obtained.
- Agitate swab briskly in vial containing viral transport media.
- Hold the end of the swab shaft; bend it at 180-degree angle to break at the marked breakpoint. Place tip in vial and secure the lid tightly.

Respiratory Site Collection Instructions

1. Throat Swab Collection:

- Use sterile **polyester-tipped swab** provided in the collection kit.
- Swab posterior oropharynx and tonsillar tissues firmly in a manner similar as for streptococcal throat culture.
- If specific lesions are seen (suggesting HSV, enterovirus, etc), use a second swab to sample the lesions.
- Agitate swab briskly in vial containing viral transport media.
- Hold the end of the swab shaft; bend it at a 180-degree angle to break at the marked breakpoint. Place tip in vial and secure the lid tightly.

2. Posterior Nasopharynx Swab Collection:

- Use sterile **flocked swab** provided in the collection kit.
- Have patient lie down on his/her back, or sit in a chair.
- Immobilize patient's head and gently insert sterile **flocked swab** into one naris while lifting the nostrils.
- Continue inserting the swab with a rotating back and forth motion until it reaches the posterior nasopharynx where resistance is met. This will require the swab to bend along the curvature of the nasopharyngeal space. The distance inserted to reach the posterior nasopharynx should be at least a few inches, depending on the age of the patient.
- Leave the swab in place for 5-10 seconds. This will likely induce tearing and coughing.
- Withdraw the swab with a rotating motion to loosen and collect cellular material while in contact with the mucosal surfaces of the mid-inferior portion of the inferior turbinate.
- Agitate swab briskly in vial containing viral transport media.
- Hold the end of the swab shaft; bend it at a 180-degree angle to break at the marked breakpoint. Place tip in vial and secure the lid tightly.



Non-Respiratory Body Site Collection Instructions

1. Anal Swab Collection:

- Wet sterile **polyester-tipped swab** with sterile saline.
- Firmly swab rectal mucosa while rotating swab.
- Agitate swab briskly in vial containing viral transport media.
- Hold the end of the swab shaft; bend it at a 180-degree angle to break at the marked breakpoint. Place tip in vial and secure the lid tightly.

2. Cervical, Vaginal, Urethral Swab Collection:

- Firmly swab desired area with sterile **polyester-tipped swab**.
- Agitate swab briskly in vial containing viral transport media.
- Hold the end of the swab shaft; bend it at a 180-degree angle to break at the marked breakpoint. Place tip in vial and secure the lid tightly.

3. Conjunctival Swab Collection:

- Firmly swab conjunctiva with sterile **polyester-tipped swab**.
- Left and right eye may be collected with **separate swabs** and pooled into one vial.
- Agitate swab briskly in vial containing viral transport media.
- Hold the end of the swab shaft; bend it at a 180-degree angle to break at the marked breakpoint. Place tip in vial and secure the lid tightly.

4. Lesion/ Vesicle Swab Collection:

- Newly developed, unbroken vesicles provide best yield.
- Break vesicle and firmly swab fluid and vesicle base (to obtain cells) with sterile **polyester-tipped swab**.
- If vesicle is already broken, select vesicle with moist base, scrape outer layer and firmly swab base.
- More than one vesicle/lesion may be sampled and the specimens pooled in one vial.
- Agitate swab briskly in vial containing viral transport media.
- Hold the end of the swab shaft; bend it at a 180-degree angle to break at the marked breakpoint. Place tip in vial and secure the lid tightly.

5. Tissue:

- Autopsy or biopsy tissue obtained for viral culture should be placed into viral transport media.

Contacting Laboratory Services

Client Services..... (614)722-5477 or (800) 934-6575

Virology Laboratory personnel can be consulted for technical and collection procedure information by calling the phone number above.