A complex rearrangement of PDGFRB elucidated by long-read sequencing



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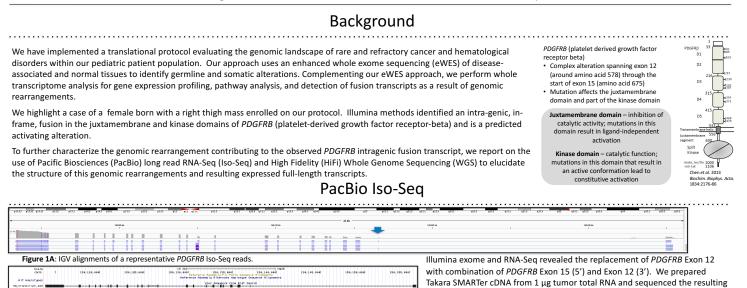




Figure 1C: Clustal Ω comparison of *PDGFRB* wild-type and rearranged amino acid sequence.

PacBio PDGFRB amplicon sequencing

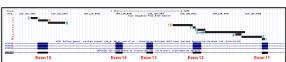
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reading frame.

spliced rearrangement transcripts.

To further examine the PDGFRB exon 12 rearrangement, PCR primers were designed to the end of PDGFRB exon 11 and the middle of PDGFRB exon 12. Amplicons were converted into SMRTbell Template Prep 1.0 libraries and sequenced on Sequel 1M. Single CCS reads were aligned to the human reference in the UCSC Genome Browser

Figure 2: UCSC Blat alignment of an annotated PacBio amplicon CCS read. The Blat alignments are numbered in the linear order of the CCS read components. The PDGFRB wild-type exon 11-intron 12 locus (716 bp) is replaced by a complex rearrangement of PDGFRB segments (1374 bp). The rearrangement contains a large, intron 11 inversion (627 bp), deletion of a portion of exon 12 (31 bp) and duplications of intron 14, exon 15 and intron 15.



SMRTbell Template Prep 1.0 library on four Sequel 1M SMRT Cells. The circular

consensus (CCS) reads were analyzed with PacBio Iso-Seq application in SMRT

Figure 1A: IGV alignments of Iso-Seq full-length wild-type and differentially

Figure 1B: UCSC Blat alignment of representative Iso-Seq transcript showing the loss of 5' Exon 12 and duplication of the 5' portion of Exon 15.. Figure 1C: Wild-type and rearrangement PDGFRB amino acid sequences were aligned with Clustal $\boldsymbol{\Omega}.$ The rearrangement transcript results in an open

Figure 2: UCSC Blat alignment of a representative PDGFRB amplicon CCS read.

PacBio WGS HiFi 15 kb circular consensus sequencing

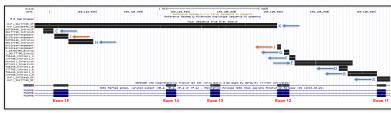
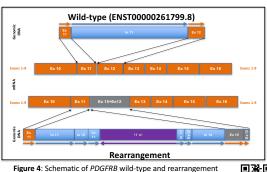


Figure 3: UCSC Blat alignment of two representative PDGFRB HiFi CCS reads

Discussion

- Genomic profiling identified a complex alteration within the PDGFRB juxtamembrane and kinase domains predicting a gain of function mutation that is therapeutically targetable with tyrosine kinase inhibitors.
- Short-read sequencing was unable to accurately resolve this complex genomic rearrangement. To elucidate the structure of this alteration, we performed PacBio long-read sequencing of normal and tumor derived DNA and the tumor derived RNA.
- PacBio Iso-Seg of the tumor RNA revealed full-length wild-type and aberrant PDGRFB transcripts. We observed RNA molecules with allele-specific isoform expression that maintain frame predicted to encode an aberrant protein products
- Long-range amplicon and long read (>15kb) whole genome sequencing (HiFi) support this somatic alteration maintaining both genomic order and orientation (Figure 4).
- The combination of rapid, cost-effective short-read sequencing and comprehensive long-read DNA and RNA sequencing provide the opportunity to diagnose and explain novel, disease-causing genetic alterations.



genomic DNA and mRNA.



All previous examinations of this tumor, either Illumina or PacBio, required enzymatic amplification. As a final exploration of this complex rearrangement, we prepared a PacBio HiFi library from 3.8 µg genomic DNA that was sheared to 15 kb and sequenced without size-fractionation using the Sequel II 8M and v2 Early Access chemistry to achieve 39X mapped sequence coverage.

Figure 3: UCSC Blat alignment of two representative PDGFRB HiFi CCS reads (19.0 and 20.1 kb). These reads encompass the PDGFRB exon 12 rearrangement as well as the start codon (in Exon 2) and the stop codon (in Exon 23). The aligned sequence, etween exon 11 and intron 15, is labeled in the order represented in the CCS reads.